




Functions of glutaminyl cyclase and its isoform in diseases

Yu'e Liu¹ , Yufeng Shi^{1,2} , and Ping Wang^{1,*}

¹ Tongji University Cancer Center, Shanghai Tenth People's Hospital of Tongji University, School of Medicine, Tongji University, Shanghai 200092, PR China

² Clinical Center for Brain and Spinal Cord Research, Tongji University, Shanghai 200092, PR China

Received 22 August 2022, Accepted 21 November 2022, Published online 6 January 2023

Abstract – Glutaminyl cyclase (QC; isoform: isoQC) is a zinc-dependent enzyme that catalyzes the intramolecular cyclization of N-terminal glutamine and glutamic acid residues into a pyroglutamate residue (pGlu). This conversion is a type of posttranslational modification called pyroglutamylation. The expression of QC/isoQC is regulated by epigenetics, cell homeostasis, and its substrates. Pyroglutamylation is an important maturation process during the synthesis and secretion of hormones, functioning in different diseases, such as Alzheimer's disease, tumors, and other kinds of chronic diseases mediated by inflammation. IsoQC has been identified as a key regulator of the CD47-SIRP α checkpoint and is critical for the pyroglutamylation of CD47 at its SIRP α binding site, thus helping cancer cells evade immune surveillance. Inhibition of isoQC blocks the interaction between CD47 and SIRP α , leading to constrained tumor growth, indicating that isoQC is a novel target for immunotherapy. Targeting isoQC overcomes the side effects of targeting CD47 because isoQC is Golgi resident and is not expressed on erythrocytes. Small molecules and antibodies have been developed to target isoQC, and some of them have been tested in preclinical or clinical studies. Here, we briefly review the discovery history of QC/isoQC and then discuss its regulation and function in different diseases, emphasizing the unique role of isoQC in immunotherapy. Finally, we summarize the development of inhibitors and their progress in clinical trials with the hope of providing useful insights for future investigation of QC/isoQC and targeting it in various diseases.

Key words: Glutaminyl cyclase, *QPCTL*, Pyroglutamylation, Phagocytosis, CD47, isoQC.

Introduction

Glutaminyl cyclase (QC) is encoded by the glutaminyl-peptide cyclotransferase (*QPCT*) gene, and the isoform of QC (isoQC) is encoded by the glutaminyl-peptide cyclotransferase-like (*QPCTL*) gene [1]. QC was first isolated from the latex of the tropical plant *Carica papaya* in 1963 [2, 3] and later from animals, plants, and bacteria [4], while the first isoQC of humans was isolated in 2008 [1]. QC is expressed in the mammalian pituitary, hypothalamus, other parts of the brain, adrenal medulla, and B lymphocytes [4, 5], with the highest expression in the striatum and hippocampus [6].

QC/isoQC catalyzes the formation of N-terminal glutamine and glutamic acid residues on target proteins into an N-terminal pyroglutamate residue (pGlu) [1]. The catalytic function of QC was first determined in plants and later in mammals [3, 4, 7]. The cyclization of the N-terminal glutamine occurs spontaneously and slowly under physiological conditions. IsoQC is highly expressed in brain tissue in Alzheimer's disease (AD)

and some chronic diseases caused by inflammation [8]. It mainly protects target proteins from degradation and plays an important role in the initiation of neurovegetative diseases and monocyte migration.

Pyroglutamate formation of the cluster of differentiation 47 (CD47) catalyzed by isoQC is critical for the binding between CD47 and the signal-regulatory protein alpha (SIRP α) at their binding site, thus contributing to cancer cell immune surveillance [9, 10]. CD47 creates a "don't eat me" signal by binding with SIRP α on myeloid cells with high specificity. Tumor cells highly express CD47 to evade immune surveillance by the CD47-SIRP α axis [11]. The CD47-SIRP α axis has been considered a tumor phagocytosis checkpoint. Targeting CD47 or interrupting the CD47-SIRP α axis exerts efficacy in solid tumors and hematological malignancies while causing adverse effects such as anemia since CD47 is also highly expressed on red blood cells (RBCs). Inhibition of isoQC enhances the phagocytosis of myeloid cells [9]. IsoQC resides in the Golgi complex, which is absent in RBCs [12–14]; therefore, targeting isoQC overcomes the adverse side effects (anemia) of targeting CD47 [15].

*Corresponding author: wangp@tongji.edu.cn

In this review, we briefly summarize the structure, regulation, and function of QC/isoQC. Then, we highlight the function of the critical enzyme isoQC in the pyroglutamylation of CD47 in the CD47-SIRP α pathway and the targeting of isoQC as a novel therapy. We believe that this short review will further our understanding of the importance of QC/isoQC in various diseases and will help researchers design a better anti-CD47 strategy in tumor immunotherapy.

Introduction and regulation of QC/isoQC

Introduction of QC/isoQC

Mammalian *QPCT* cDNA was isolated from the bovine pituitary in 1991 [6]. The gene encodes a glycoprotein QC containing sulfhydryl groups [4], a zinc-binding site [16, 17], and a mixed α -helix and β -sheet structure functioning in the polarization of the γ -amide group of the substrate and the interaction with competitive inhibitors of QC [18].

QCs exhibit overlapping but distinct homologies, structures, and characteristics among different species [6, 19–22]. The murine *QPCT* gene is located on chromosome 17, and the human *QPCT* is on chromosome 2; the murine *QPCTL* is located on chromosome 7, while the human *QPCTL* is on chromosome 19. Both *QPCT* and its isoform *QPCTL* are ubiquitously expressed in mice; *QPCT* expression is higher in neuronal tissue, while there is no significant difference in *QPCTL* expression in tissues [1]. Human *QPCTL* and murine *QPCTL* display broad substrate specificity and a preference for hydrophobic substrates [23].

As a metalloenzyme, both human QC and isoQC contain a zinc-binding domain, which reveals the conservation of the zinc-binding motif [1]. Human isoQC carries an N-terminal signal anchor that is different from that in QC [1]. Human isoQC exhibits 10-fold lower activity than human QC, indicating that isoQC has an overall lower enzymatic activity [1]. Human QC colocalizes with the Golgi apparatus, endoplasmic reticulum, and secretory granules, where it is thought to play an important role in the maturation of different proteins [1, 24], while its isoform is exclusively localized within the Golgi complex and is obviously retained by the N-terminus [1]. IsoQC shows 46% sequence identity to QC, but they have almost identical substrate specificity *in vitro* [1].

Regulation of QC/isoQC expression

The expression of QC/isoQC is tightly regulated at multiple levels, including epigenetic mechanisms, cell homeostasis, and its substrates.

Histone methylation activates QC maternally

QPCTL has been predicted to be a potential placenta-specific imprinted gene expressed maternally. Genomic imprinting is an epigenetic process by which the male and female germ lines confer specific marks (imprints) onto certain gene regions, leading to one allele with the imprinted gene being active and the other one being silenced [25–27]. QC could

be a target for the treatment of diseases related to the development of the placenta and embryo. The DNA methylation level does not affect the imprinting level of QC, but histone methylation of H3Kme3 results in maternal activation of QC [28]. Whether isoQC is also regulated by H3Kme3 has not been explored and needs further investigation.

Ca²⁺ homeostasis enhances QC expression and its enzyme activity

Ca²⁺ homeostasis enhances QC mRNA expression and enzyme activity. The QC promoter contains a putative binding site for the calcium-dependent transcription factors *c-fos* and *c-jun*, which are induced by Ca²⁺-related stimuli, and their upregulation enhances QC expression. In neuronal cells, the selective upregulation of QC via Ca²⁺-dependent transcription factors is the major consequence of unsettled Ca²⁺ homeostasis [29]. Whether Ca²⁺ homeostasis regulates the expression of isoQC is not clear.

Proinflammatory cytokines upregulate QC/isoQC under inflammation

Chemokines play a pivotal role in different inflammatory disorders, such as atherosclerosis fibrosis, AD, and tumor progression [30–32]. The expression of QC/isoQC is upregulated by proinflammatory cytokines such as CCL2 and CX3C chemokine ligand 1 (CX3CL1), which act as its substrates.

CCL2 is a chemokine known to recruit monocytes to sites of inflammation, playing important roles in cancer progression [33]. CX3CL1 is a multifunctional inflammatory chemokine whose function depends on several factors, including its structures [34]. Both CCL2 and CX3CL1 are typical cytokines that mediate cell adhesion and migration in inflammatory processes [35], and both are substrates of QC/isoQC. The N-terminus of both chemokines has a glutamine in the first position and can be post-translationally modified by QC/isoQC to form a pGlu residue [35]. Under inflammatory conditions, the release of chemokines such as CCL2 and CX3CL1 increases the expression of QC/isoQC at the mRNA level, thus ensuring that all these chemokines are released in their fully pyroglutamylated forms [35]. Furthermore, the expression of QC/isoQC correlates with CCL2 expression at the mRNA level in an NF- κ B-dependent manner [1, 33].

The function of QC/isoQC

QC/isoQC is a zinc-dependent enzyme that catalyzes the intramolecular cyclization of N-terminal glutamine and glutamic acid residues into pGlu [36]. This conversion is a type of posttranslational modification named pyroglutamylation. Pyroglutamylation occurs at the N-terminus of a number of peptide hormones and secretory proteins, such as gonadotropin-releasing hormone (GnRH), thyrotropin-releasing hormone (TRH), neurotensin and fibronectin [37, 38].

IsoQC possesses almost the same substrates as QC but differs in subcellular localization [1, 23, 39]. The functions of QC and isoQC probably complement each other, suggesting

a pivotal role of pyroglutamylation in protein and peptide maturation [23].

QC/isoQC function in protein stabilization

The formation of the N-terminal pGlu is an important maturation step during the synthesis and secretion of hormones [40, 41]. The pyroglutamylation modification allows regulatory peptides to adopt a proper conformation for binding to their receptors and renders the protein more resistant to protease degradation [4, 7] and more susceptible to hydrophobic interactions, aggregation, and neurotoxicity [7, 42, 43]. In snakes, QC and isoQC also contribute to the stabilization of peptide toxins [44].

QC/isoQC protects proteins in cerebrospinal fluid from degradation

Proteins and peptides in human cerebrospinal fluid (CSF) are potential substrates of QC/isoQC, whose activity is a characteristic feature of CSF [43]. The pyroglutamylation of proteins in CSF stabilizes the peptide from degradation by aminopeptidases [45]. This may play an important role in neurological disorders, which are characterized by the inappropriate expression of QCs or their substrates [46].

QC/isoQC function in chemokine activity and stability

The pyroglutamylation of CCL2 catalyzed by both QC and isoQC plays critical roles in the synthesis, secretion, maturation, and function of CCL2 [1, 39, 47] and protects CCL2 from proteolytic degradation during these processes *in vivo* [39, 48]. The activity and stability of CCL2 also depend on its pyroglutamylation [1, 39, 47].

QC/isoQC also catalyzes the N-terminal pyroglutamylation of CX3CL1, which is important for its stability and its interaction with the CX3CL1 receptor and provides new insights into the function of QC in inflammation [35]. Loss of pyroglutamylation results in increased accessibility for N-terminal proteolytic degradation or directly affects ligand-receptor interaction (Fig. 1a).

QC/isoQC function in monocyte migration

QC-catalyzed pyroglutamylation of monocyte chemoattractant proteins is required for monocyte migration in the inflammatory process; thus, QC may be a potential target for some inflammatory disorders [48].

QC/isoQC plays essential roles in monocyte homeostasis. It supports mouse monocyte migration and sustains monocyte infiltration and tumor growth. Pharmacological isoQC inhibition prevents monocyte accumulation in established tumors and promotes the remodeling of macrophage compartments. Suppressing isoQC also improves CD8⁺ T-cell responses upon programmed cell death-ligand 1 (PD-L1) blockade and enhances antitumor activity. Moreover, isoQC inhibition constitutes an effective approach for myeloid cell-targeted cancer immunotherapy, and disruption of isoQC can be used in

combination with checkpoint inhibitors to enhance the antibody-mediated phagocytosis of tumor cells [49].

The function of isoQC in cancer immunotherapy

Brief introduction of CD47

CD47 is a glycosylated 5-transmembrane protein expressed on many types of cells. It is composed of a glycosylated N-terminal extracellular domain (ECD), a 5-transmembrane-spanning domain, and a short C-terminal domain (CTD) [50–53]. The ECD contains a V-set immunoglobulin superfamily (IgSF) domain and is a cell surface marker of self that binds to SIRP α .

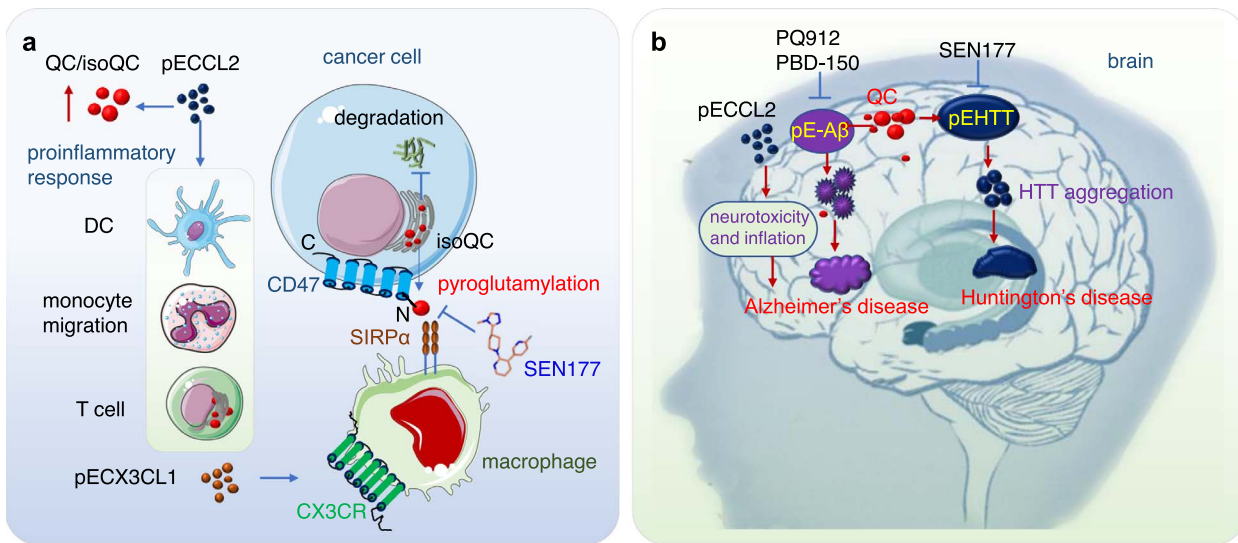
The expression of CD47 is regulated at the transcriptional level, translational level, and posttranslational level. Transcription factors such as Myc [54, 55], hypoxia-inducible factor-1 (HIF-1) α [56–58], The nuclear factor κ B (NF- κ B) [59, 60], and α -Pal/NRF-1 [61, 62] regulate the expression of CD47; various cytokines such as tumor necrosis factor (TNF)- α [59, 63, 64], interferon- γ [65, 66], and interleukins [67, 68] potentiate CD47 expression. Posttranscriptional modification of CD47, including glycosylation and pyroglutamylation, regulates CD47 function [69, 70]. The glycosylation of CD47 is essential for its binding to thrombospondin 1 (TSP-1) [71, 72], and the pyroglutamylation of CD47 is essential for the CD47-SIRP α axis to function in immunotherapy [9, 10].

CD47 plays important roles not only in biological processes such as proliferation, autophagy, and phagocytosis but also in various types of cancer. It has been regarded as a key antiphagocytic molecule that renders tumor cells resistant to immune surveillance.

The CD47-SIRP α pathway

CD47 regulates a cascade of signaling pathways by binding to its receptors, such as SIRP α and TSP-1. SIRP α is a transmembrane glycoprotein that contains three Ig-like domains, one transmembrane domain, and four tyrosine phosphorylation sites with two immune receptor tyrosine-based inhibitor motifs (ITIMs) in the cytoplasmic tail [73–75]. The NH₂-terminal domain of SIRP α binds to the IgV domain of CD47, and the interaction between CD47 and SIRP α leads to the phosphorylation of ITIMs in SIRP α . Phosphorylated ITIMs recruit and activate the inhibitory molecules Src homology 2 (SH2) domain-containing protein tyrosine phosphatase (SHP)-1 and SHP-2. The recruitment of SHP-1 and SHP-2 enables the phosphorylation of myosin IIA and then suppresses the function of non-muscle myosin IIA, which modulates phagolysosomal biogenesis in macrophages and plays important roles in phagocytosis [76]. Finally, the interaction between CD47 and SIRP α inhibits phagocytosis and cytotoxicity [76–78]. When the binding between CD47 and SIRP α is blocked, the latter's ITIMs are not phosphorylated, and the lack of SHP-1 and SHP-2 recruitment enables the activation of prophagocytic receptors to trigger phagocytosis.

CD47 creates a “don't eat me” signal by binding SIRP α on myeloid cells with high specificity. The CD47-SIRP α axis inhibits phagocytosis and protects RBCs from



QC/isoQC function in cancer

QC/isoQC function in neuron diseases

Figure 1. The function of QC/isoQC in diseases. (a) *Function of QC/isoQC in cancer:* IsoQC catalyzes the pyroglutamylation of CD47 and contributes to the interaction with the CD47-SIRP α axis and finally leads to the cancer cell immune surveillance evasion. Targeting isoQC by SEN177 or other molecules inhibits the pyroglutamylation of CD47, thus CD47 cannot interact with SIRP α , and cancer cells are phagocytosed by the macrophages. The pyroglutamylation of CCL2 upregulates CCL2 expression and recruits more immune cells such as DCs, monocytes, and T cells into the tumor microenvironment. The pyroglutamylation of CX3CL1 is critical for its binding to CX3CR and blocks the CX3CL pyroglutamylation leading to the degradation of the protein and affecting its binding to receptors. The basic function of QC/isoQC is to catalyze the target protein N terminal pyroglutamine and protect it from degradation. (b) *Function of QC in neuro diseases:* QC is primarily and highly expressed in the brain including the hippocampus and cortex and other peripheral tissues. It converts beta-amyloid (A β) N-terminal peptides at position 3 or 11 into pGlu. Abnormally upregulation of QC in AD functions as a key inducer in the initiation of AD by catalyzing the generation of different mediators such as pyroglutamylation of A β and CCL2, which are the main causes of neurotoxicity and inflammation. Small molecules such as PBD-150 and PQ-912 inhibit QC activity and reduce the syndrome of AD, PQ-912 is in a Phase II clinical trial now. In Huntington's disease, the pyroglutamylation catalyzed by QCs leads to the aggregation of huntingtin (HTT) protein and finally leads to Huntington's disease. A small molecule such as SEN177 targeting QC/isoQC rescues Huntington's disease phenotypes efficiently and reduce HTT protein aggregation.

erythrophagocytosis [79]. Tumor cells utilize this mechanism to evade immune surveillance by highly expressing CD47 [11, 80–85]. Therefore, the CD47-SIRP α axis has been considered a tumor phagocytosis checkpoint [83–86]. Targeting CD47 or disrupting the CD47-SIRP α axis has shown efficacy against solid tumors and hematological malignancies while causing adverse effects such as anemia since CD47 is also highly expressed on RBCs. Therefore, it is important to gain a deeper understanding of the CD47-SIRP α axis with the hope of identifying modulators that could be targeted for myeloid cell checkpoint blockade.

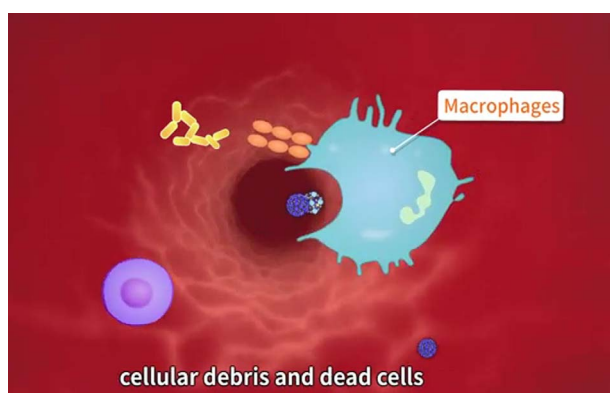
The pyroglutamylation of CD47 contributes to the CD47-SIRP α pathway

Crystal structures of the CD47-SIRP α interaction have revealed the involvement of pyroglutamate at the N-terminus of CD47 based on its hydrogen bonding [87]. The N-terminal Q19 of CD47 is modified to a pGlu residue by isoQC [10]. This process involves the loss of an NH $_3$ from N-terminal glutamine, leading to the formation of a cyclic amide structure, thus generating the pyroglutamate derivative [43]. IsoQC-mediated CD47 modification occurs very early in the protein life cycle

[9]. This pyroglutamate is assumed to form spontaneously upon cleavage of the CD47 signal sequence [88]. The role of isoQC as a modifier of CD47 is observed in many cells, including malignant melanoma and other types of cancer cells [9].

The N-terminal pyroglutamylation of CD47 is an important posttranslational modification of CD47, which can be specifically recognized by SIRP α and contributes to the interaction between CD47 and SIRP α [9, 87]. CD47-SIRP α binding critically depends on a CD47 posttranslational modification of its N-terminal glutamine to a pyroglutamate [9]. isoQC was discovered to be involved in CD47-SIRP α by two different screens in two labs. Logtenberg et al. used a fluorescence-activated cell sorting (FACS)-based haploid genetic screen with an antibody against human CD47 and found that isoQC is a typical CD47 regulator [9]. Our lab used a FACS-based whole-genome screen in the colon cancer cell line HCT116 and obtained similar results [10]. By coculturing tumor cells and macrophages, we found that isoQC inhibits the phagocytosis of tumor cells by macrophages.

The interaction between CD47 and SIRP α is druggable by compounds such as QC/isoQC inhibitors, which have been verified by two independent studies [9, 10]. This indicates that pyroglutamylation serves as an immunotherapeutic target by



Video 1. <https://vcm.edpsciences.org/10.1051/vcm/2022008#V1>.

abolishing immune checkpoint signaling and that isoQC is a potential therapeutic target for the treatment of various cancers (Video 1).

QC/isoQC function in diseases

QC and isoQC play important roles in cancers such as melanoma [89], papillary thyroid carcinoma [90, 91], and renal cell carcinoma [28, 92]. Abnormal upregulation of isoQC in AD functions as a key inducer in the initiation of AD, catalyzing the generation of different mediators, such as pyroglutamylation of A β s (pE-A β s) and pE-CCL2, which are the main causes of neurotoxicity and inflammation [93–99]. More details on QC/isoQC function in diseases will be addressed in the next section.

Targeting QC/isoQC in cancers and other diseases

Targeting QC/isoQC in cancers

QC/isoQC correlates with survival and drug resistance

QC/isoQC is highly expressed in melanoma [100, 101], papillary thyroid carcinoma [90], glioma, cholangiocarcinoma, and colon cancer. The relationship of QC/isoQC expression with patient survival is inconsistent. In most cancers, the high expression of QC/isoQC is associated with a lower survival rate, while in cholangiocarcinoma and colon adenocarcinoma, the high expression of QC/isoQC indicates a higher survival rate [102]. In sunitinib-nonresponsive renal cell carcinoma (RCC), the methylation level in QC is decreased, suggesting QC may function in tumor progression and drug resistance [103]. QC colocalizes with HRAS in the cytoplasm and binds HRAS to promote its stability and reduce its ubiquitin-mediated degradation, finally leading to drug resistance [103]. Therefore, the expression of QC may be related to sensitivity to drugs in RCC.

Targeting isoQC in the CD47-SIRP α pathway in cancers

CD47-SIRP α is the key pathway through which tumor cells escape phagocytosis by macrophages, and pyroglutamylation of CD47 is the key modification for CD47 to bind to SIRP α [104].

IsoQC is the central enzyme catalyzing the pyroglutamylation of CD47. Pyroglutamylation contributes to the macrophage-mediated phagocytosis of tumor cells [9, 10]. Genetic and pharmacological interference with isoQC activity enhances antibody-dependent cellular phagocytosis and increases the cellular cytotoxicity of tumor cells and the neutrophil-mediated killing of tumor cells *in vivo* [9, 10]. Knockout of isoQC induces the accumulation of macrophages expressing phagocytic genes, indicating that this modality of antitumor immunity may be enhanced by two isoQC-mediated mechanisms. Targeting isoQC significantly reduces the binding ability of CD47 to SIRP α to regulate tumor immunity [10, 105, 106]. Unlike CD47-targeted monoclonal antibodies, isoQC inhibitors avoid the side effects on RBCs. In addition, a recent article reported that isoQC depletion also results in impaired monocyte recruitment, affected monocyte homeostasis, and improved CD8⁺ T-cell responses [49] (Fig. 2).

The facilitating function of isoQC in the CD47-SIRP α pathway and in monocyte migration suggests that targeting isoQC is a means of engaging and enhancing multiple antitumor mechanisms, including macrophage-mediated phagocytosis and adaptive T-cell responses.

Targeting QC/isoQC in other diseases

QC/isoQC plays important roles in the pathology of several diseases, such as rheumatoid arthritis, osteoporosis [107], AD [107, 108], and septic arthritis [109]. IsoQC is significantly upregulated and closely related to decreased patient survival [10].

QC/isoQC in inflammatory diseases

Acute and chronic inflammatory disorders are characterized by detrimental cytokine and chemokine expression. The chemotactic activity of cytokines depends on a modified N-terminus of the polypeptide. For example, CCL2 is modified by pyroglutamylation to protect it against degradation *in vivo*. QC/isoQC inhibitors attenuate atherosclerotic pathology in a model of accelerated atherosclerosis. A combinatorial approach of QC/isoQC knockout as well as the pharmacological inhibition of QC/isoQC represents an alternative therapeutic strategy to treat CCL2-driven disorders such as atherosclerosis/restenosis and fibrosis [39]. Targeting QC/isoQC activity leads to the degradation of CCL2 and the reduction of kidney inflammation [110].

Nonalcoholic fatty liver disease (NAFLD) is the most prevalent form of hepatic pathology, and inflammation is an integral part of NAFLD. The chemokine CCL2 and its primary receptor CCR2 are key regulators of inflammation. The pyroglutamylation of CCL2 protects it from degradation by aminopeptidases, and this truncated form reduces its potential to attract immune cells in the inflamed liver [111]. Therefore, inhibition of QC/isoQC is a promising therapy for NAFLD.

QC/isoQC in neuron diseases

(a) IsoQC in Alzheimer's disease

AD is a common chronic and progressive neurodegenerative disease [112]. Deposition of amyloid- β (A β) remains a

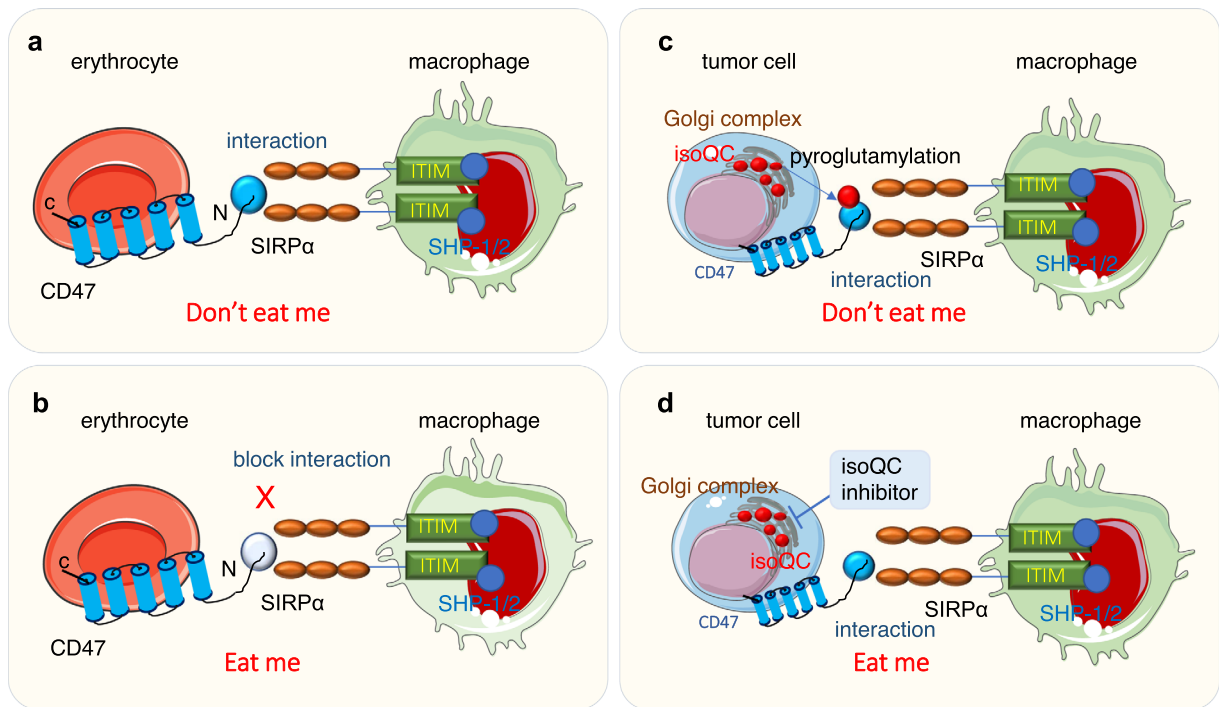


Figure 2. Targeting isoQC in cancer immunotherapy. The interaction between CD47 on erythrocyte and SIRP α on macrophages triggers a “don’t eat me” signal, thus the erythrocyte is not engulfed by macrophages. (b) Interrupting the CD47-SIRP α pathway either by blocking CD47 or SIRP α lead to the phagocytosis of erythrocytes by macrophages. (c) The interaction between CD47 on tumor cells and SIRP α on macrophages triggers a “don’t eat me” signal, thus the tumor cells are not engulfed by macrophages, by this mechanism, tumor cells realize their immune surveillance evasion. The pyroglutamylation of CD47 is critical for the binding between CD47 and SIRP α . (d) Blocking the CD47-SIRP α pathway leads to the phagocytosis of tumor cells by macrophages but meanwhile brings the adverse effect that the erythrocytes are phagocytized by the macrophages. Targeting isoQC by SEN177 or other inhibitors inhibits the pyroglutamylation of CD47 and thus blocks the interaction between CD47 and SIRP α . IsoQC is a Golgi-resident enzyme that is not expressed on erythrocytes, thus the erythrocytes would not be affected by targeting isoQC.

hallmark feature of AD, yet the precise mechanisms by which this peptide induces neurotoxicity remain unknown. Research on this point has grown exponentially over the past few decades. The contribution of pyroglutamylation of A β to the neurodegenerative process in AD has been widely studied over the last few years [113, 114].

QC is primarily and highly expressed in the brain, including the hippocampus and cortex, as well as in peripheral tissues. It is highly involved in the posttranslational modification of A β peptides by converting its N-terminal peptide at position 3 or 11 into pGlu. Abnormal upregulation of QC in AD functions as a key inducer in the initiation of AD by catalyzing the generation of different mediators, such as pyroglutamylation of A β and CCL2, which are the main causes of neurotoxicity and inflammation [93–99] (Fig. 1b).

Pyroglutamylated A β peptides (A β pEs) are found in self-aggregated A β peptides and act as initiators of A β accumulation, favoring plaque seeding [115, 116]. A β pE is only found in AD brains and constitutes approximately 50% of the total A β [117], so the level of A β pE has been treated as a potential biomarker of AD [118]. A β pE is upstream of the neurotoxic amyloid cascade, triggering neurodegeneration and enhancing AD pathology [119–121].

A β pE has been treated as a drug target in AD, and drugs such as donanemab (a humanized IgG1 monoclonal antibody

to the pyroglutamylated form of A β) are in phase II clinical trials [122–126]. Many A β peptides possess a pGlu residue catalyzed by QC [127], and the pyroglutamylation of A β is highly abundant in the brains of AD patients, protecting it from protease degradation and making it more hydrophobic and prone to aggregation, which might increase its neurotoxicity [24, 101, 128, 129]. QC catalyzes the pyroglutamylation of A β , and QC inhibition lowers the amount of pyroglutamylated A β and lessens the AD-like pathology. Oral application of a QC inhibitor resulted in reduced A β (3pE-42) burden in two different transgenic mouse models of AD and in a new *Drosophila* model [101].

In conclusion, QC/isoQC is a potential target in drug development due to its pronounced stability, neurotoxicity, and aggregation propensity, and various inhibitors have been synthesized targeting pyroglutamylation in AD [130].

(b) QC in Huntington’s disease

Huntington’s disease (HD) is an incurable neurodegenerative disease caused by an abnormally expanded polyglutamine tract in the huntingtin protein (HTT) [131]. In a cell-based siRNA screen, QC had one of the strongest effects on mutant HTT-induced toxicity and aggregation, and it rescued these phenotypes in *Drosophila*. Targeting QC/isoQC with new inhibitors using *in silico* methods followed by *in vitro* screening

rescued the phenotype and disrupted HTT aggregation [131] (Fig. 1b).

QC/isoQC in osteoporosis

Osteoporosis is a disease of low bone mass, high bone fragility, and high morbidity [132]. Hormone levels are involved in the occurrence and progression of osteoporosis [133]. QC/isoQC is an essential modifier of pituitary peptide hormones, including GnRH, which is associated with osteoporosis. Genetic variations in QC are important factors affecting the bone mineral density (BMD) of adult women and thereby contribute to susceptibility to osteoporosis [107].

The common polymorphisms in QC are closely associated with BMD in both the Chinese and Japanese populations [107, 134]. The *QPCT* gene lies on chromosome 2p22.3, within a region influencing forearm BMD, suggesting bone mass differences between individuals may be influenced by the nucleotide variations of the *QPCT* gene. By investigating 13 known polymorphisms, it was found that a single-nucleotide polymorphism (SNP) variation at isoQC contributes to BMD in adult women [107].

In conclusion, the expression and function of QC/isoQC in many types of diseases are under research, and advanced research is focused on neuron-related diseases. Its function and working mechanisms in cancer require more investigation. Due to its importance in various diseases, targeting QC/isoQC by small molecules is developing fast.

Small molecules targeting QC/isoQC

Since QC and isoQC have 46% conserved sequences, including sites with catalytic activity, many QC substrates can also be catalyzed by isoQC to form pyroglutamide, suggesting that they have similar substrate selectivity [1]. Theoretically, inhibitors of QC would also inhibit isoQC, as demonstrated by Holger et al. [1]. IsoQC, as an isoenzyme, is an important target to reduce the risk of cross-reactivity and side effects.

QC inhibitors developed from 2004 to 2021 have been reviewed elsewhere [135, 136]. Many inhibitors of QC have been prepared and screened *in vitro* [16, 18, 101, 137–145]. Since the catalytic activity of QC is zinc-dependent, some types of QC inhibitors were designed based on its zinc domain, such as imidazole, triazole, and benzimidazole [146], and they all contain, in general, the following distinct structural features: a zinc-binding group, a hydrogen-bonding donor and an aromatic group to interact with Phe325 in the pocket as a key amino acid for potent binding. They mimic two or three amino acids of the sequence of *N*-truncated A β , NH₂–Glu–Phe–Arg [18, 137–139, 147].

Imidazole and its derivatives were identified as competitive inhibitors of QC. They interact with accessible zinc atoms in the active site of carboxypeptidase A to form stable complexes with active-site residues of serine proteases in the presence of zinc ions [148–150]. The first effective inhibitors for human QC were described in 2006 [17, 18]. Most of the inhibitors of QC/isoQC were designed based on the imidazole/methylimidazole ring as a zinc-binding motif (motif 1). Later, more compounds presented extended motifs, consisting of an

H-bonding donor (motif 2) and an aromatic moiety (motif 3) or even a fourth motif mimicking an arginine guanidinium moiety [18, 137, 140–142]. The minimal requirements to obtain a high-affinity inhibitor are a Zn (II)-binding moiety, either imidazole or triazole and a hydrophobic aromatic ring [151]. Some compounds are predicted to be able to permeate the blood-brain barrier (BBB), and they can be used as oral drugs [152]. Here, we summarize a couple of QC and isoQC inhibitors.

PBD-150

Researchers have further modified imidazole to improve its inhibition of QC enzyme activity [23] and found that thiourea derivative modification on the nitrogen atom of imidazole No. 1 can greatly improve its inhibition of QC [39].

PBD-150 is a human QC Y115E–Y117E variant inhibitor with a high affinity for QC (inhibition constant $K_i = 490$ nM for human QC and $K_i = 173$ nM for murine QC) [18, 153]. PBD-150 thiourea is a representative compound. Its inhibition of QC is more potent than that of isoQC (isoQC, $K_i = 7.3$ μ M; QC, $K_i = 60$ nM) [128, 154]. PBD-150 is the prototype QC inhibitor with imidazole as a zinc-binding group, and it was discovered by Probiodrug AG through fragment-based screening [18].

PBD-150 can reduce the deposition and aggregation of pyroglutamate-modified amyloid- β peptides in the brains of transgenic mouse models of AD, leading to a significant improvement in learning and memory in those transgenic animals [18, 101, 155]. Furthermore, PBD150 plays important role in monocyte migration simulated by monocyte chemoattractant proteins; it reduces N-terminal uncyclized-MCP-stimulated monocyte migration without affecting pGlu-containing MCP-induced cell migration [48].

Even PBD-150 has been demonstrated *in vitro* and *in vivo* to decrease the formation of pGlu-A β , but the preclinical evaluation of PBD-150 demonstrates that it cannot cross the BBB [154] due to the polarity of this compound [156], suggesting that the therapeutic effect of QC inhibition may be associated with peripheral mechanisms [156]. Future works in the development of this therapeutic agent may focus on the improvement of BBB permeability, which would let it serve as a companion diagnostic when developed as a PET radioligand.

SEN177

The SEN177-binding mode to human QC differs from that of the known human QC inhibitors. SEN177 is designed based on a triazine ring as the human QC-binding motif and brings different extension motifs [151]. Crystal studies of the SEN177–human QC complex show two interactions between the enzyme and the inhibitor in terms of inhibitor orientation: the pyridine ring with Trp207 and the fluorine atom of 2-fluoropyridine with the hydrogen atom of His330 [152]. SEN177 exhibits comparable or better potency compared to other inhibitors, such as PBD-150 and PQ912 ($K_i = 24.7$ nM) [151]. Its K_i to human QC is 20 nM, and its half-maximum inhibitory concentration is only 0.013 μ M for isoQC, so it is one of the best inhibitors of isoQC.

SEN177 is the first reported small-molecule antagonist of the SIRP α -CD47 interaction. Treatment with the isoQC inhibitor SEN177 reduces human SIRP α -Fc staining in many cancer cell lines, thus leading to the reduced binding of CD47-SIRP α even though the total CD47 levels are not significantly affected [9]. Knockout of isoQC reduces the binding of the CD47 antibody CC2C6, which has the same recognition site (pGlu) as SIRP α [9]. The SIRP α -CD47 interaction is also quantitatively measured in live and fixed tumor cells via a new assay relying on laser scanning cytometry (LSC) [4]. Furthermore, blocking CD47-SIRP α by SEN177 enhances tumor cell killing by polymorphonuclear neutrophilic granulocytes (PMNs) with different EGFR antibody isotypes [157].

In addition, SEN177 can efficiently rescue HD phenotypes and reduce HTT protein aggregation in mammalian cell lines, mouse neurons, *Drosophila* eyes, and zebrafish [131].

PQ529

PQ529 is a newly reported isoform-nonspecific QC/isoQC inhibitor [39]; its full name is 1-(1H-benzo[d]imidazol-5-yl)-5-(4-propoxyphenyl)imidazolidine-2,4-dione, and it possesses benzimidazole as a warhead. The K_i -values of PQ529 at pH 8 are 38 nM for human QC, 4 nM for human isoQC, 27 nM for murine QC, and 2 nM for murine isoQC [109]. Treatment with PQ529 results in a significant delay in the development of arthritis [109]. PQ529 significantly inhibits CCL2 and amyloid β *in vivo* and *in vitro*. It has potential therapeutic effects on atherosclerosis and AD [39, 158].

PQ912

PQ912 (drug name: Varoglutamstat) is an inhibitor of QC that plays a critical role in the formation of synaptotoxic pyroglutamate-A-beta oligomers (A β Os) and has no significant toxicities when used in neurodegenerative diseases. The model of action of PQ912 is as follows: A β Os, which are synaptotoxic and lead to cognitive decline, play important roles in the pathophysiology of AD [159–161]. pE-A β s initiate and sustain the formation of synaptotoxic A β Os. PQ912 is a QC inhibitor that can decrease the formation of pE-A β [162]. Therefore, inhibition of QC with PQ912 leads to a decrease in pE-A β s and consequentially a reduction in A β Os [115]. Treatment with PQ912 in a mouse model of AD improves cognition and attenuates the pathology of AD [144, 163]. PQ912 prevents the formation of pGlu3-A β in different compartments of cells, and the combination of PQ912 and a pGlu3-A β -specific antibody (m6) in transgenic mice exert additive effects on brain A β pathology [164].

PQ912 is a first-in-class QC inhibitor in clinical development, and it has undergone rigorous preclinical and clinical investigations, including *in vitro* and *in vivo* animal and phase 1 clinical studies [144, 162]. A phase 2a study aims to investigate the highest dose of PQ912 that was used in a phase I study of the same compound to evaluate its safety and tolerability in patients with mild cognitive impairment due to AD. The results have shown that PQ912 is safe and highly effective [162, 165]. It is currently under phase 2b study, and the results are expected in early 2023 [166].

In addition to all the above inhibitors, new heterocyclic and peptidomimetic derivatives to inhibit QC and isoQC are still under investigation. In Dec. 2021, the cyclopentylmethyl derivative (214) exhibited the most potent *in vitro* activity (IC₅₀ = 0.1 nM), which is 290-fold higher than that of PQ912, and benzimidazole (227) showed the most promising *in vivo* efficacy [123].

Natural products targeting QC/isoQC

In addition to small molecules, natural products and traditional Chinese medicines are widely used for various diseases [167]. A myriad of isolated natural products, such as natural phenol compounds, marine products, and flavonoid derivatives, have revealed new active scaffolds with isoQC-inhibitory properties, and some of them have exhibited promising and exciting results [130, 140].

Flavonoids exhibit many desired pharmacological effects, including antioxidation and anti-inflammation [168, 169]. Luteolin, a natural flavonoid compound, has potent anticancer effects *in vitro* and *in vivo* due to its natural antioxidant ability [170, 171]. Luteolin has been verified as a novel inhibitor of isoQC by our lab; it directly interacts with isoQC protein, reduces pGlu modification of CD47 and its surface binding to SIRP α , and finally promotes macrophage-mediated phagocytosis [172]. Other natural compounds from microalgae have been synthesized to target QC or isoQC [173].

Conclusion and prospects

QC/isoQC is a critical enzyme that functions in both biological and pathological conditions via the pyroglutamylation of proteins. Most of the research focuses on its function in AD or other neurodegenerative diseases. We explored and summarized its function in immunotherapy due to its unique role in the CD47-SIRP α checkpoint.

Cancer immunotherapy is now a pillar of cancer treatment that continues to develop. Cancer immunotherapies, including checkpoint inhibitors and adoptive cell therapy, manipulate the immune system to recognize and attack cancer cells [174]. Cancer cells escape immune surveillance by hijacking inhibitory pathways through the overexpression of checkpoint genes; thus, the downregulation of checkpoint gene expression can be a potential weapon in immunotherapy. Besides T cells, macrophages and neutrophils are also targets of immunotherapy [9, 175]. IsoQC blocking is a novel immunotherapy approach that relies on innate myeloid checkpoint (CD47-SIRP α) blockade [176, 177]. The physiological role of isoQC is probably to stabilize bioactive peptides, and the unique role of isoQC in macrophage-mediated phagocytosis gives it a role in cancer immunotherapy. In CD47-highly-expressing tumors, targeting CD47 suppresses tumor growth but also has side effects, such as limited penetration of tumor tissues associated with their structure and large size, which may impede its clinical application. Targeting isoQC provides new opportunities to increase the therapeutic efficacy of tumor-directed antibodies by interfering with the CD47-SIRP α axis. Targeting isoQC not only avoids the anemia problem but also leads to better tumor penetration, fewer infusion reactions, and higher patient compliance [178].

Knowledge of QC/isoQC has increased considerably, and the therapeutic potential of QC/isoQC inhibitors has been explored. Difficulties in obtaining large quantities of pure protein may limit the use of crystallographic screening for drug development on this target, and the complicated metabolism *in vivo* requires more experiments [179, 180]. QC enzymes are pharmacologically interesting targets to be used as an AD-modifying therapy [136]. Many inhibitors have been developed and optimized, but only PQ-912 has been applied in clinical trials and has completed a phase 2a trial in AD [165]. Regarding the application of isoQC targeting in cancer immunotherapy, no inhibitors are on the market until now. In Feb. 2022, Insilico Medicine nominated the preclinical candidate compound ISM004-1057D, which targets isoQC in the CD47-SIRP α pathway, for innovative tumor immunotherapy. ISM004-1057D is in the investigational new drug (IND) stage. Once the clinical trial application is approved, the cooperating company Fosun Pharma will conduct clinical trials immediately.

In the future, innate myeloid checkpoint blockade will become another important therapeutic option [176, 177], and isoQC as a potential target deserves more investigation.

Conflict of interest

The authors declare that they have no conflict of interest.

Acknowledgements. This work was supported by grants from the National Natural Science Foundation of China (Nos. 31830053, 31920103007, 8207112072), the National Key Research and Development Program of China (2020YFA0803201), the Science Technology Commission of Shanghai Municipality (No. 20S11900700).

Author contributions

P.W. designed the study. Y.L. contributed to the literature search, designed the video, and wrote the paper. J.Z. gave suggestions in revision. Y.S. and P.W. revised the manuscript based on their rich research experience. All authors read and approved the final manuscript.

References

- Cynis H, Rahfeld JU, Stephan A, et al. Isolation of an isoenzyme of human glutaminyl cyclase: retention in the Golgi complex suggests involvement in the protein maturation machinery. *Journal of Molecular Biology*. 2008;379(5):966–980.
- Azarkan M, Wintjens R, Looze Y, et al. Detection of three wound-induced proteins in papaya latex. *Phytochemistry*. 2004;65(5):525–534.
- Messer M. Enzymatic cyclization of L-glutamine and L-glutaminyl peptides. *Nature*. 1963;197:1299.
- Busby WH, Quackenbush GE, Humm J, et al. An enzyme(s) that converts glutaminyl-peptides into pyroglutaminyl-peptides – presence in pituitary, brain, adrenal-medulla, and lymphocytes. *Journal of Biological Chemistry*. 1987;262(18):8532–8536.
- Vale W, Spiess J, Rivier C, et al. Characterization of a 41-residue ovine hypothalamic peptide that stimulates secretion of corticotropin and beta-endorphin. *Science*. 1981;213(4514):1394–1397.
- Pohl T, Zimmer M, Mugele K, et al. Primary structure and functional expression of a glutaminyl cyclase. *Proceedings of the National Academy of Sciences of the United States of America*. 1991;88(22):10059–10063.
- Fischer WH, Spiess J. Identification of a mammalian glutaminyl cyclase converting glutaminyl into pyroglutaminyl peptides. *Proceedings of the National Academy of Sciences of the United States of America*. 1987;84(11):3628–3632.
- Wang XJ, Wang L, Yu X, et al. Glutaminyl cyclase inhibitor exhibits anti-inflammatory effects in both AD and LPS-induced inflammatory model mice. *International Immunopharmacology*. 2019;75:105770.
- Logtenberg MEW, Jansen JHM, Raaben M, et al. Glutaminyl cyclase is an enzymatic modifier of the CD47-SIRP alpha axis and a target for cancer immunotherapy. *Nature Medicine*. 2019;25(4):612–619.
- Wu ZQ, Weng LJ, Zhang TB, et al. Identification of glutaminyl cyclase isoenzyme isoQC as a regulator of SIRP alpha-CD47 axis. *Cell Research* 2019;29(6):502–505.
- Zhao XW, van Beek EM, Schornagel K, et al. CD47-signal regulatory protein-alpha (SIRPalpha) interactions form a barrier for antibody-mediated tumor cell destruction. *Proc Natl Acad Sci U S A*. 2011;108(45):18342–18347.
- Moras M, Lefevre SD, Ostuni MA. From erythroblasts to mature red blood cells: organelle clearance in mammals. *Frontiers in Physiology*. 2017;8:1076.
- Ch R, Rey G, Ray S, et al. Rhythmic glucose metabolism regulates the redox circadian clockwork in human red blood cells. *Nature Communications*. 2021;12(1):377.
- Wiback SJ, Palsson BO. Extreme pathway analysis of human red blood cell metabolism. *Biophysical Journal*. 2002;83(2):808–818.
- Ingram JR, Blomberg OS, Sockolosky JT, et al. Localized CD47 blockade enhances immunotherapy for murine melanoma. *Proc Natl Acad Sci U S A*. 2017;114(38):10184–10189.
- Schilling S, Cynis H, von Bohlen A, et al. Isolation, catalytic properties, and competitive inhibitors of the zinc-dependent murine glutaminyl cyclase. *Biochemistry*. 2005;44(40):13415–13424.
- Schilling S, Niestroj AJ, Rahfeld JU, et al. Identification of human glutaminyl cyclase as a metalloenzyme – Potent inhibition by imidazole derivatives and heterocyclic chelators. *Journal of Biological Chemistry*. 2003;278(50):49773–49779.
- Buchholz M, Heiser U, Schilling S, et al. The first potent inhibitors for human glutaminyl cyclase: synthesis and structure-activity relationship. *Journal of Medicinal Chemistry*. 2006;49(2):664–677.
- Bockers TM, Kreutz MR, Pohl T. Glutaminyl-cyclase expression in the bovine/porcine hypothalamus and pituitary. *Journal of Neuroendocrinology*. 1995;7(6):445–453.
- Oberg KA, Ruyschaert JM, Azarkan M, et al. Papaya glutamine cyclase, a plant enzyme highly resistant to proteolysis, adopts an all-beta conformation. *European Journal of Biochemistry*. 1998;258(1):214–222.
- Wintjens R, Belrhali H, Clantin B, et al. Crystal structure of papaya glutaminyl cyclase, an archetype for plant and bacterial glutaminyl cyclases. *Journal of Molecular Biology*. 2006;357(2):457–470.
- Alam M, Ho S, Vance DE, et al. Heterologous expression, purification, and characterization of human triacylglycerol hydro-lase. *Protein Expression of Purification*. 2002;24(1):33–42.
- Stephan A, Wermann M, von Bohlen A, et al. Mammalian glutaminyl cyclases and their isoenzymes have identical enzymatic characteristics. *FEBS Journal*. 2009;276(22):6522–6536.
- Hartlage-Rubsamen M, Staffa K, Waniek A, et al. Developmental expression and subcellular localization of glutaminyl

- cyclase in mouse brain. *International Journal of Developmental Neuroscience*. 2009;27(8):825–835.
25. Reik W, Walter J Genomic imprinting: parental influence on the genome. *Nature Reviews Genetics*. 2001;2(1):21–32.
 26. Horsthemke B, Mechanisms of imprint dysregulation. *American Journal of Medical Genetics Part C Seminars in Medical Genetics*. 2010;154C(3):321–328.
 27. Liu Y, Chen C, Wang X, et al. An epigenetic role of mitochondria in cancer. *Cells*. 2022;11(16):2518.
 28. Guo J, He H, Liu Q, et al. Identification and epigenetic analysis of a maternally imprinted gene Qpct. *Molecular and Cellular*. 2015;38(10):859–865.
 29. De Kimpe L, Bennis A, Zwart R, et al. Disturbed Ca²⁺ homeostasis increases glutaminyl cyclase expression; connecting two early pathogenic events in Alzheimer's disease in vitro. *PLoS One*. 2012;79.
 30. Charo IF, Taubman MB Chemokines in the pathogenesis of vascular disease. *Circulation Research*. 2004;95(9):858–866.
 31. Inoshima I, Kuwano K, Hamada N, et al. Anti-monocyte chemoattractant protein-1 gene therapy attenuates pulmonary fibrosis in mice. *American Journal of Physiology-Lung Cellular and Molecular Physiology*. 2004;286(5):L1038–L1044.
 32. Galimberti D, Fenoglio C, Lovati C, et al. Serum MCP-1 levels are increased in mild cognitive impairment and mild Alzheimer's disease. *Neurobiology of Aging*. 2006;27(12):1763–1768.
 33. Kehlen A, Haegele M, Menge K, et al. Role of glutaminyl cyclases in thyroid carcinomas. *Endocrine-Related Cancer*. 2013;20(1):79–90.
 34. Conroy MJ, Lysaght J CX3CL1 signaling in the tumor microenvironment. *Advances in Experimental Medicine and Biology*. 2020;1231:1–12.
 35. Kehlen A, Haegele M, Bohme L, et al. N-terminal pyroglutamate formation in CX3CL1 is essential for its full biologic activity. *Bioscience Reports*. 2017;37(4):BSR20170712.
 36. Schilling S, Manhart S, Hoffmann T, et al. Substrate specificity of glutaminyl cyclases from plants and animals. *Biological Chemistry*. 2003;384(12):1583–1592.
 37. Awade AC, Cleuziat P, Gonzales T, et al. Pyrrolidone carboxyl peptidase (Pcp) – an enzyme that removes pyroglutamic acid (pGlu) from pGlu-peptides and pGlu-proteins. *Proteins-Structure Function and Bioinformatics*. 1994;20(1):34–51.
 38. Blombäck B [44] Derivatives of glutamine in peptides. In: *Methods in Enzymology*. Academic Press;1967. p. 398–411.
 39. Cynis H, Hoffmann T, Friedrich D, et al. The isoenzyme of glutaminyl cyclase is an important regulator of monocyte infiltration under inflammatory conditions. *EMBO Molecular Medicine*. 2011;3(9):545–558.
 40. Goren HJ, Bauce LG, Vale W, Forces and structural limitations of binding of thyrotrophin-releasing factor to the thyrotrophin-releasing receptor: the pyroglutamic acid moiety. *Molecular Pharmacology*. 1977;13(4):606–614.
 41. Abraham GN, Podell DN, Pyroglutamic acid. Non-metabolic formation, function in proteins and peptides, and characteristics of the enzymes effecting its removal. *Molecular and Cellular Biochemistry*. 1981;38(Spec No(Pt 1)):181–190.
 42. Wang Z, Sun B, Zhu F, Molecular characterization of glutaminyl-peptide cyclotransferase(QPCT)in *Scylla paramamosain* and its role in *Vibrio alginolyticus* and white spot syndrome virus (WSSV) infection. *Fish and Shellfish Immunology*. 2018;78:299–309.
 43. Schilling S, Hoffmann T, Manhart S, et al. Glutaminyl cyclases unfold glutamyl cyclase activity under mild acid conditions. *FEBS Letters*. 2004;563(1–3):191–196.
 44. Pawlak J, Manjunatha Kini R, Snake venom glutaminyl cyclase. *Toxicon*. 2006;48(3):278–286.
 45. Benter IF, Hirsh EM, Tuchman AJ, et al. N-terminal degradation of low molecular weight opioid peptides in human cerebrospinal fluid. *Biochemical Pharmacology*. 1990;40(3):465–472.
 46. Gontsarova A, Kaufmann E, Tuman H, et al. Glutaminyl cyclase activity is a characteristic feature of human cerebrospinal fluid. *Clinica Chimica Acta*. 2008;389(1–2):152–159.
 47. Van Coillie E, Proost P, Van Aelst I, et al. Functional comparison of two human monocyte chemotactic protein-2 isoforms, role of the amino-terminal pyroglutamic acid and processing by CD26/dipeptidyl peptidase IV. *Biochemistry*. 1998;37(36):12672–12680.
 48. Chen YL, Huang KF, Kuo WC, et al. Inhibition of glutaminyl cyclase attenuates cell migration modulated by monocyte chemoattractant proteins. *Biochemical Journal*. 2012;442(2):403–412.
 49. Barreira da Silva R, Leitao RM, Pechuan-Jorge X, et al. Loss of the intracellular enzyme QPCTL limits chemokine function and reshapes myeloid infiltration to augment tumor immunity. *Nature Immunology*. 2022;23(4):568–580.
 50. Matlung HL, Szilagyi K, Barclay NA, et al. The CD47-SIRP alpha signaling axis as an innate immune checkpoint in cancer. *Immunological Reviews*. 2017;276(1):145–164.
 51. Reinhold MI, Lindberg FP, Plas D, et al. In vivo expression of alternatively spliced forms of integrin-associated protein (CD47). *Journal of Cell Science*. 1995;108(Pt 11):3419–3425.
 52. Lindberg FP, Gresham HD, Schwarz E, et al. Molecular cloning of integrin-associated protein: an immunoglobulin family member with multiple membrane-spanning domains implicated in alpha v beta 3-dependent ligand binding. *Journal of Cell Biology*. 1993;123(2):485–496.
 53. Oldenborg PA, Zheleznyak A, Fang YF, et al. Role of CD47 as a marker of self on red blood cells. *Science*. 2000;288(5473):2051–2054.
 54. Casey SC, Tong L, Li Y, et al. MYC regulates the antitumor immune response through CD47 and PD-L1. *Science*. 2016;352(6282):227–231.
 55. Li W, Gupta SK, Han W, et al. Targeting MYC activity in double-hit lymphoma with MYC and BCL2 and/or BCL6 rearrangements with epigenetic bromodomain inhibitors. *Journal of Hematology & Oncology*. 2019;12(1):73.
 56. Zhang H, Lu H, Xiang L, et al. HIF-1 regulates CD47 expression in breast cancer cells to promote evasion of phagocytosis and maintenance of cancer stem cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2015;112(45):E6215–E6223.
 57. Monti E, Marras E, Prini P, et al. Luteolin impairs hypoxia adaptation and progression in human breast and colon cancer cells. *European Journal of Pharmacology*. 2020;881:173210.
 58. Samanta D, Park Y, Ni X, et al. Chemotherapy induces enrichment of CD₄₇⁺/CD₇₃⁺/PDL₁⁺ immune evasive triple-negative breast cancer cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2018;115(6):E1239–E1248.
 59. Lo J, Lau EY, Ching RH, et al. Nuclear factor kappa B-mediated CD47 up-regulation promotes sorafenib resistance and its blockade synergizes the effect of sorafenib in hepatocellular carcinoma in mice. *Hepatology*. 2015;62(2):534–545.
 60. Candas-Green D, Xie B, Huang J, et al. Dual blockade of CD47 and HER2 eliminates radioresistant breast cancer cells. *Nature Communications*. 2020;11(1):4591.

61. Virbasius CA, Virbasius JV, Scarpulla RC NRF-1, an activator involved in nuclear-mitochondrial interactions, utilizes a new DNA-binding domain conserved in a family of developmental regulators. *Genes & Development*. 1993;7(12A):2431–2445.
62. Gomezcuadrado A, Martin M, Noel M, et al. Initiation binding-receptor, a factor that binds to the transcription initiation site of the histone h5 gene, is a glycosylated member of a family of cell-growth regulators. *Molecular and Cellular Biology*. 1995;15(12):6670–6685.
63. Betancur PA, Abraham BJ, Yiu YY, et al. A CD47-associated super-enhancer links pro-inflammatory signalling to CD47 upregulation in breast cancer. *Nature Communications*. 2017; 8:14802.
64. Zhang X, Wang Y, Fan J, et al. Blocking CD47 efficiently potentiated therapeutic effects of anti-angiogenic therapy in non-small cell lung cancer. *Journal for ImmunoTherapy of Cancer*. 2019;7(1):346.
65. Ye ZH, Jiang XM, Huang MY, et al. Regulation of CD47 expression by interferon-gamma in cancer cells. *Translational Oncology*. 2021;14(9):101162.
66. Sockolosky JT, Dougan M, Ingram JR, et al. Durable antitumor responses to CD47 blockade require adaptive immune stimulation. *Proceedings of the National Academy of Sciences of the United States of America*. 2016;113(19):E2646–E2654.
67. Chen J, Zheng DX, Yu XJ, et al. Macrophages induce CD47 upregulation via IL-6 and correlate with poor survival in hepatocellular carcinoma patients. *Oncoimmunology*. 2019;8(11):e1652540.
68. Bian Z, Shi L, Guo YL, et al. Cd47-Sirpalha interaction and IL-10 constrain inflammation-induced macrophage phagocytosis of healthy self-cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2016;113(37):E5434–E5443.
69. Shi R, Chai Y, Duan X, et al. The identification of a CD47-blocking “hotspot” and design of a CD47/PD-L1 dual-specific antibody with limited hemagglutination. *Signal Transduction and Targeted Therapy*. 2020;5(1):16.
70. Parthasarathy R, Subramanian S, Boder ET, et al. Post-translational regulation of expression and conformation of an immunoglobulin domain in yeast surface display. *Biotechnology and Bioengineering*. 2006;93(1):159–168.
71. Isenberg JS, Annis DS, Pendrak ML, et al. Differential interactions of thrombospondin-1, -2, and -4 with CD47 and effects on cGMP signaling and ischemic injury responses. *Journal of Biological Chemistry*. 2009;284(2):1116–1125.
72. Kaur S, Kuznetsova SA, Pendrak ML, et al. Heparan sulfate modification of the transmembrane receptor CD47 is necessary for inhibition of T cell receptor signaling by thrombospondin-1. *Journal of Biological Chemistry*. 2011;286(17):14991–15002.
73. Brooke GP, Parsons KR, Howard CJ, Cloning of two members of the SIRP alpha family of protein tyrosine phosphatase binding proteins in cattle that are expressed on monocytes and a subpopulation of dendritic cells and which mediate binding to CD4 T cells. *European Journal of Immunology*. 1998;28(1):1–11.
74. Barclay AN, van den Berg TK, The interaction between signal regulatory protein alpha (SIRP alpha) and CD47: structure, function, and therapeutic target. *Annual Review of Immunology*. 2014;32(32):25–50.
75. Fujioka Y, Matozaki T, Noguchi T, et al. A novel membrane glycoprotein, SHPS-1, that binds the SH2-domain-containing protein tyrosine phosphatase SHP-2 in response to mitogens and cell adhesion. *Molecular and Cellular Biology*. 1996;16(12):6887–6899.
76. Tsai RK, Discher DE, Inhibition of “self” engulfment through deactivation of myosin-II at the phagocytic synapse between human cells. *Journal of Cell Biology*. 2008;180(5):989–1003.
77. Okazawa H, Motegi S, Ohyama N, et al. Negative regulation of phagocytosis in macrophages by the CD47-SHPS-1 system. *Journal of Immunology*. 2005;174(4):2004–2011.
78. Matozaki T, Murata Y, Okazawa H, et al. Functions and molecular mechanisms of the CD47-SIRPalpha signalling pathway. *Trends in Cell Biology*. 2009;19(2):72–80.
79. Huang CY, Ye ZH, Huang MY, et al. Regulation of CD47 expression in cancer cells. *Translational Oncology*. 2020;13(12):100862.
80. Chao MP, Alizadeh AA, Tang C, et al. Anti-CD47 antibody synergizes with rituximab to promote phagocytosis and eradicate non-Hodgkin lymphoma. *Cell*. 2010;142(5):699–713.
81. Chao MP, Alizadeh AA, Tang C, et al. Therapeutic antibody targeting of CD47 eliminates human acute lymphoblastic leukemia. *Cancer Research*. 2011;71(4):1374–1384.
82. Rendtew Danielsen JM, Knudsen LM, Dahl IM, et al. Dysregulation of CD47 and the ligands thrombospondin 1 and 2 in multiple myeloma. *British Journal of Haematology*. 2007; 138(6):756–760.
83. Chan KS, Espinosa I, Chao M, et al. Identification, molecular characterization, clinical prognosis, and therapeutic targeting of human bladder tumor-initiating cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2009;106(33):14016–14021.
84. Yoshida K, Tsujimoto H, Matsumura K, et al. CD47 is an adverse prognostic factor and a therapeutic target in gastric cancer. *Cancer Medicine*. 2015;4(9):1322–1333.
85. Willingham SB, Volkmer JP, Gentles AJ, et al. The CD47-signal regulatory protein alpha (SIRPα) interaction is a therapeutic target for human solid tumors. *Proceedings of the National Academy of Sciences of the United States of America*. 2012;109(17):6662–6667.
86. Gao AG, Lindberg FP, Finn MB, et al. Integrin-associated protein is a receptor for the C-terminal domain of thrombospondin. *Journal of Biological Chemistry*. 1996;271(1):21–24.
87. Hatherley D, Graham SC, Turner J, et al. Paired receptor specificity explained by structures of signal regulatory proteins alone and complexed with CD47. *Molecular Cell*. 2008;31(2):266–277.
88. Ho CCM, Guo N, Sockolosky JT, et al. “Velcro” engineering of high affinity CD47 ectodomain as signal regulatory protein alpha (SIRP alpha) antagonists that enhance antibody-dependent cellular phagocytosis. *Journal of Biological Chemistry*. 2015;290(20):12650–12663.
89. Gillis JS Microarray evidence of glutaminyl cyclase gene expression in melanoma: implications for tumor antigen specific immunotherapy. *Journal of Translational Medicine*. 2006; 4:27.
90. da Silveira Mitteldorf CA, de Sousa-Canavez JM, Leite KR, et al. FN1, GALE, MET, and QPCT overexpression in papillary thyroid carcinoma: molecular analysis using frozen tissue and routine fine-needle aspiration biopsy samples. *Diagnostic Cytopathology*. 2011;39(8):556–561.
91. Jarzab B, Wiensch M, Fujarewicz K, et al. Gene expression profile of papillary thyroid cancer: sources of variability and diagnostic implications. *Cancer Research*. 2005;65(4):1587–1597.
92. Morris MR, Ricketts CJ, Gentle D, et al. Genome-wide methylation analysis identifies epigenetically inactivated candidate tumour suppressor genes in renal cell carcinoma. *Oncogene*. 2011;30(12):1390–1401.

93. Hartlage-Rubsamen M, Morawski M, Waniek A, et al. Glutaminyl cyclase contributes to the formation of focal and diffuse pyroglutamate (pGlu)-Abeta deposits in hippocampus via distinct cellular mechanisms. *Acta Neuropathologica*. 2011; 121(6):705–719.
94. Morawski M, Schilling S, Kreuzberger M, et al. Glutaminyl cyclase in human cortex: correlation with (pGlu)-amyloid-beta load and cognitive decline in Alzheimer's disease. *Journal of Alzheimer's Disease*. 2014;39(2):385–400.
95. Schilling S, Lauber T, Schaupp M, et al. On the seeding and oligomerization of pGlu-amyloid peptides (in vitro). *Biochemistry*. 2006;45(41):12393–12399.
96. D'Arrigo C, Tabaton M, Perico A, N-terminal truncated pyroglutamate beta amyloid peptide A beta py3-42 shows a faster aggregation kinetics than the full-length A beta 1–42. *Biopolymers*. 2009;91(10):861–873.
97. Jawhar S, Wirths O, Bayer TA, Pyroglutamate amyloid-beta (Abeta): a hatchet man in Alzheimer disease. *Journal of Biological Chemistry*. 2011;286(45):38825–38832.
98. Bridel C, Hoffmann T, Meyer A, et al. Glutaminyl cyclase activity correlates with levels of Abeta peptides and mediators of angiogenesis in cerebrospinal fluid of Alzheimer's disease patients. *Alzheimer's Research & Therapy*. 2017;9(1):38.
99. Hartlage-Rubsamen M, Waniek A, Meissner J, et al. Isoglutaminyl cyclase contributes to CCL2-driven neuroinflammation in Alzheimer's disease. *Acta Neuropathologica*. 2015;129(4): 565–583.
100. Muthusamy V, Duraisamy S, Bradbury CM, et al. Epigenetic silencing of novel tumor suppressors in malignant melanoma. *Cancer Research*. 2006;66(23):11187–11193.
101. Schilling S, Zeitschel U, Hoffmann T, et al. Glutaminyl cyclase inhibition attenuates pyroglutamate Abeta and Alzheimer's disease-like pathology. *Nature Medicine*. 2008;14(10): 1106–1111.
102. Tang Z, Kang B, Li C, et al. GEPIA2: an enhanced web server for large-scale expression profiling and interactive analysis. *Nucleic Acids Research*. 2019;47(W1):W556–W560.
103. Zhao T, Bao Y, Gan X, et al. DNA methylation-regulated QPCT promotes sunitinib resistance by increasing HRAS stability in renal cell carcinoma. *Theranostics*. 2019;9(21): 6175–6190.
104. Mateo V, Lagneaux L, Bron D, et al. CD47 ligation induces caspase-independent cell death in chronic lymphocytic leukemia. *Nature Medicine*. 1999;5(11):1277–1284.
105. Mair B, Aldridge PM, Atwal RS, et al. High-throughput genome-wide phenotypic screening via immunomagnetic cell sorting. *Nature Biomedical Engineering*. 2019;3(10):796–805.
106. Logtenberg MEW, Glutaminyl cyclase is an enzymatic modifier of the CD47-SIRP alpha axis and target for immunotherapy. *Cancer Immunology Research*. 2019;7(2).
107. Ezura Y, Kajita M, Ishida R, et al. Association of multiple nucleotide variations in the pituitary glutaminyl cyclase gene (QPCT) with low radial BMD in adult women. *Journal of Bone and Mineral Metabolism*. 2004;19(8):1296–1301.
108. Batliwalla FM, Baechler EC, Xiao X, et al. Peripheral blood gene expression profiling in rheumatoid arthritis. *Genes & Immunity*. 2005;6(5):388–397.
109. Hellvard A, Maresz K, Schilling S, et al. Glutaminyl cyclases as novel targets for the treatment of septic arthritis. *Journal of Infectious Diseases*. 2013;207(5):768–777.
110. Kanemitsu N, Kiyonaga F, Mizukami K, et al. Chronic treatment with the (iso-)glutaminyl cyclase inhibitor PQ529 is a novel and effective approach for glomerulonephritis in chronic kidney disease. *Naunyn-Schmiedeberg's Archives of Pharmacology*. 2021;394(4):751–761.
111. Cynis H, Kehlen A, Haegele M, et al. Inhibition of glutaminyl cyclases alleviates CCL2-mediated inflammation of non-alcoholic fatty liver disease in mice. *International Journal of Experimental Pathology*. 2013;94(3):217–225.
112. Minter MR, Taylor JM, Crack PJ, The contribution of neuroinflammation to amyloid toxicity in Alzheimer's disease. *Journal of Neurochemistry*. 2016;136(3):457–474.
113. Thal DR, Walter J, Saïdo TC, et al. Neuropathology and biochemistry of Abeta and its aggregates in Alzheimer's disease. *Acta Neuropathologica*. 2015;129(2):167–182.
114. Gunn AP, Wong BX, McLean C, et al. Increased glutaminyl cyclase activity in brains of Alzheimer's disease individuals. *Journal of Neurochemistry*. 2021;156(6):979–987.
115. Nussbaum JM, Schilling S, Cynis H, et al. Prion-like behaviour and tau-dependent cytotoxicity of pyroglutamylated amyloid-beta. *Nature*. 2012;485(7400):651–655.
116. Schlenzig D, Manhart S, Cinar Y, et al. Pyroglutamate formation influences solubility and amyloidogenicity of amyloid peptides. *Biochemistry*. 2009;48(29):7072–7078.
117. Kuo YM, Emmerling MR, Woods AS, et al. Isolation, chemical characterization, and quantitation of A beta 3-pyroglutamyl peptide from neuritic plaques and vascular amyloid deposits. *Biochemical and Biophysical Research Communications*. 1997;237(1):188–191.
118. Wu HQ, Can small molecule inhibitors of glutaminyl cyclase be used as a therapeutic for Alzheimer's disease?. *Future Medicinal Chemistry*. 2017;9(17):1979–1981.
119. Pivtoraiko VN, Abrahamson EE, Leurgans SE, et al. Cortical pyroglutamate amyloid-beta levels and cognitive decline in Alzheimer's disease. *Neurobiol Aging*. 2015;36(1):12–19.
120. Upadhya AR, Kosterin I, Kumar S, et al. Biochemical stages of amyloid-beta peptide aggregation and accumulation in the human brain and their association with symptomatic and pathologically preclinical Alzheimer's disease. *Brain*. 2014; 137:887–903.
121. Moro ML, Phillips AS, Gaimster K, et al. Pyroglutamate and isoaspartate modified amyloid-Beta in ageing and Alzheimer's disease. *Acta Neuropathologica Communications*. 2018;6(1):3.
122. Mintun MA, Lo AC, Duggan Evans C, et al. Donanemab in early Alzheimer's disease. *New England Journal of Medicine*. 2021;384(18):1691–1704.
123. Van Manh N, Hoang VH, Ngo VTH, et al. Discovery of highly potent human glutaminyl cyclase (QC) inhibitors as anti-Alzheimer's agents by the combination of pharmacophore-based and structure-based design. *European Journal of Medicinal Chemistry*. 2021;226:113819.
124. Bayer TA, Wirths O Focusing the amyloid cascade hypothesis on N-truncated Abeta peptides as drug targets against Alzheimer's disease. *Acta Neuropathologica*. 2014;127(6): 787–801.
125. DeMattos Ronald B., Lu J, Tang Y, et al. A plaque-specific antibody clears existing β -amyloid plaques in Alzheimer's disease mice. *Neuron*. 2012;76(5):908–920.
126. Lowe SL, Willis BA, Hawdon A, et al. Donanemab (LY3002813) dose-escalation study in Alzheimer's disease. *Alzheimer's & Dementia (New York, NY)*. 2021;7(1):e12112.
127. Gunn AP, Masters CL, Cherny RA, Pyroglutamate-Abeta: Role in the natural history of Alzheimer's disease. *International Journal of Biochemistry & Cell Biology*. 2010;42(12): 1915–1918.
128. Cynis H, Scheel E, Saïdo TC, et al. Amyloidogenic processing of amyloid precursor protein: evidence of a pivotal role of

- glutaminy cyclase in generation of pyroglutamate-modified amyloid-beta. *Biochemistry*. 2008;47(28):7405–7413.
129. Saido TC, Iwatsubo T, Mann DM, et al. Dominant and differential deposition of distinct beta-amyloid peptide species, A beta N3(pE), in senile plaques. *Neuron*. 1995;14(2):457–466.
 130. Coimbra JR, Sobral PJ, Santos AE, et al. An overview of glutaminy cyclase inhibitors for Alzheimer's disease. *Future Medicinal Chemistry*. 2019;11(24):3179–3194.
 131. Jimenez-Sanchez M, Lam W, Hannus M, et al. siRNA screen identifies QPCT as a druggable target for Huntington's disease. *Nature Chemical Biology*. 2015;11(5):347–354.
 132. Srivastava M, Deal C, Osteoporosis in elderly: prevention and treatment. *Clinics in Geriatric Medicine*. 2002;18(3): 529–555.
 133. Gosset A, Pouilles JM, Tremollieres F Menopausal hormone therapy for the management of osteoporosis. *Best Practice & Research Clinical Endocrinology & Metabolism*. 2021;35(6): 101551.
 134. Huang QY, Kung AWC, The association of common polymorphisms in the QPCT gene with bone mineral density in the Chinese population. *Journal of Human Genetics*. 2007; 52(9):757–762.
 135. Xu C, Wang YN, Wu H, Glutaminy cyclase, diseases, and development of glutaminy cyclase inhibitors. *Journal of Medicinal Chemistry*. 2021;64(10):6549–6565.
 136. Coimbra JRM, Salvador JAR, A patent review of glutaminy cyclase inhibitors (2004-present). *Expert Opinion on Therapeutic Patents*. 2021;31(9):809–836.
 137. Buchholz M, Hamann A, Aust S, et al. Inhibitors for human glutaminy cyclase by structure based design and bioisosteric replacement. *Journal of Medicinal Chemistry*. 2009;52(22): 7069–7080.
 138. Ramsbeck D, Buchholz M, Koch B, et al. Structure-activity relationships of benzimidazole-based glutaminy cyclase inhibitors featuring a heteroaryl scaffold. *Journal of Medicinal Chemistry*. 2013;56(17):6613–6625.
 139. Tran PT, Hoang VH, Thorat SA, et al. Structure-activity relationship of human glutaminy cyclase inhibitors having an N-(5-methyl-1H-imidazol-1-yl)propyl thiourea template. *Bioorganic & Medicinal Chemistry*. 2013;21(13):3821–3830.
 140. Li M, Dong Y, Yu X, et al. Inhibitory effect of flavonoids on human glutaminy cyclase. *Bioorganic & Medicinal Chemistry*. 2016;24(10):2280–2286.
 141. Hoang VH, Tran PT, Cui M, et al. Discovery of potent human glutaminy cyclase inhibitors as anti-Alzheimer's agents based on rational design. *Journal of Medicinal Chemistry*. 2017;60(6):2573–2590.
 142. Li M, Dong Y, Yu X, et al. Synthesis and evaluation of diphenyl conjugated imidazole derivatives as potential glutaminy cyclase inhibitors for treatment of Alzheimer's disease. *Journal of Medicinal Chemistry*. 2017;60(15):6664–6677.
 143. Ngo VTH, Hoang VH, Tran PT, et al. Potent human glutaminy cyclase inhibitors as potential anti-Alzheimer's agents: Structure-activity relationship study of Arg-mimetic region. *Bioorganic & Medicinal Chemistry*. 2018;26(5):1035–1049.
 144. Hoffmann T, Meyer A, Heiser U, et al. Glutaminy cyclase inhibitor PQ912 improves cognition in mouse models of Alzheimer's disease-studies on relation to effective target occupancy. *Journal of Pharmacology and Experimental Therapeutics*. 2017;362(1):119–130.
 145. Manh NV, Hoang VH, Ngo VTH, et al. Discovery of potent indazole-based human glutaminy cyclase (QC) inhibitors as Anti-Alzheimer's disease agents. *European Journal of Medicinal Chemistry*. 2022;244.
 146. Gulcan HO, Mavideniz A, Sahin MF, et al. Benzimidazole-derived compounds designed for different targets of Alzheimer's disease. *Current Medicinal Chemistry*. 2019;26(18): 3260–3278.
 147. Ngo VTH, Hoang VH, Tran PT, et al. Structure-activity relationship investigation of Phe-Arg mimetic region of human glutaminy cyclase inhibitors. *Bioorganic & Medicinal Chemistry*. 2018;26(12):3133–3144.
 148. Lee KJ, Joo KC, Kim EJ, et al. A new type of carboxypeptidase a inhibitors designed using an imidazole as a zinc coordinating ligand. *Bioorganic & Medicinal Chemistry*. 1997;5(10):1989–1998.
 149. Katz BA, Clark JM, Finer-Moore JS, et al. Design of potent selective zinc-mediated serine protease inhibitors. *Nature*. 1998;391(6667):608–612.
 150. Dhanak D, Burton G, Christmann LT, et al. Metal mediated protease inhibition: Design and synthesis of inhibitors of the human cytomegalovirus (hCMV) protease. *Bioorganic & Medicinal Chemistry Letters*. 2000;10(20):2279–2282.
 151. Pozzi C, Di Pisa F, Benvenuti M, et al. The structure of the human glutaminy cyclase-SEN177 complex indicates routes for developing new potent inhibitors as possible agents for the treatment of neurological disorders. *Journal of Biological Inorganic Chemistry*. 2018;23(8):1219–1226.
 152. Tran PT, Hoang VH, Lee J, et al. In vitro and in silico determination of glutaminy cyclase inhibitors. *RSC Advances*. 2019;9(51):29619–29627.
 153. DiPisa F, Pozzi C, Benvenuti M, et al. The soluble Y115E–Y117E variant of human glutaminy cyclase is a valid target for X-ray and NMR screening of inhibitors against Alzheimer disease. *Acta Crystallographica Section F-Structural Biology Communications*. 2015;71:986–992.
 154. Jackson I, Brooks A, Shao X, et al. Preclinical evaluation of [¹¹C]PBD150, a glutaminy cyclase inhibitor for the detection of Alzheimer's disease prior to amyloid β burden. *Journal of Nuclear Medicine*. 2015;56(Supplement 3):1096.
 155. Huang KF, Liaw SS, Huang WL, et al. Structures of human Golgi-resident glutaminy cyclase and its complexes with inhibitors reveal a large loop movement upon inhibitor binding. *Journal of Biological Chemistry*. 2011;286(14): 12439–12449.
 156. Brooks AF, Jackson IM, Shao X, et al. Synthesis and evaluation of [¹¹C]PBD150, a radiolabeled glutaminy cyclase inhibitor for the potential detection of Alzheimer's disease prior to amyloid beta aggregation. *MedChemComm*. 2015;6(6): 1065–1068.
 157. Baumann N, Rosner T, Jansen JHM, et al. Enhancement of epidermal growth factor receptor antibody tumor immunotherapy by glutaminy cyclase inhibition to interfere with CD47/signal regulatory protein alpha interactions. *Cancer Science*. 2021;112(8):3029–3040.
 158. Cynis H, Funkelstein L, Toneff T, et al. Pyroglutamate-amyloid-beta and glutaminy cyclase are colocalized with amyloid-beta in secretory vesicles and undergo activity-dependent, regulated secretion. *Neurodegenerative Diseases*. 2014;14(2):85–97.
 159. Selkoe DJ, Alzheimer's disease is a synaptic failure. *Science*. 2002;298(5594):789–791.
 160. Selkoe DJ, Hardy J, The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Molecular Medicine*. 2016;8(6): 595–608.
 161. Ferreira ST, Lourenco MV, Oliveira MM, et al. Soluble amyloid-beta oligomers as synaptotoxins leading to cognitive

- impairment in Alzheimer's disease. *Frontiers in Cellular Neuroscience*. 2015;9:191.
162. Lues I, Weber F, Meyer A, et al. A phase 1 study to evaluate the safety and pharmacokinetics of PQ912, a glutaminy cyclase inhibitor, in healthy subjects. *Alzheimer's & Dementia (New York, NY)*. 2015;1(3):182–195.
163. Briels CT, Stam CJ, Scheltens P, et al. In pursuit of a sensitive EEG functional connectivity outcome measure for clinical trials in Alzheimer's disease. *Clinical Neurophysiology*. 2020; 131(1):88–95.
164. Hoffmann T, Rahfeld JU, Schenk M, et al. Combination of the glutaminy cyclase inhibitor PQ912 (varoglutamstat) and the murine monoclonal antibody PBD-C06 (m6) shows additive effects on brain Abeta pathology in transgenic mice, *International Journal of Molecular Sciences*. 2021;22(21): 11791.
165. Scheltens P, Hallikainen M, Grimmer T, et al. Safety, tolerability and efficacy of the glutaminy cyclase inhibitor PQ912 in Alzheimer's disease: results of a randomized, double-blind, placebo-controlled phase 2a study. *Alzheimer's Research & Therapy*. 2018;10(1):107.
166. Vijverberg EGB, Axelsen TM, Bihlet AR, et al. Rationale and study design of a randomized, placebo-controlled, double-blind phase 2b trial to evaluate efficacy, safety, and tolerability of an oral glutaminy cyclase inhibitor varoglutamstat (PQ912) in study participants with MCI and mild AD-VIVIAD. *Alzheimer's Research & Therapy*. 2021;13(1):142.
167. Huang S, Liu Y, Zhang Y, et al. Baicalein inhibits SARS-CoV-2/VSV replication with interfering mitochondrial oxidative phosphorylation in a mPTP dependent manner. *Signal Transduction and Targeted Therapy*. 2020;5(1):266.
168. Cho JG, Song NY, Nam TG, et al. Flavonoids from the grains of C1/R-S transgenic rice, the transgenic oryza sativa spp. japonica, and their radical scavenging activities. *Journal of Agricultural and Food Chemistry*. 2013;61(43):10354–10359.
169. Nijveldt RJ, van Nood E, van Hooft DE, et al. Flavonoids: a review of probable mechanisms of action and potential applications. *American Journal of Clinical Nutrition*. 2001;74(4): 418–425.
170. Devi KP, Rajavel T, Nabavi SF, et al. Hesperidin: A promising anticancer agent from nature. *Industrial Crops and Products*. 2015;76:582–589.
171. Imran M, Rauf A, Abu-Izneid T, et al. Luteolin, a flavonoid, as an anticancer agent: A review (vol. 112, 108612, 2019). *Biomedicine & Pharmacotherapy*. 2019;116:109084.
172. Li Z, Gu X, Rao D, et al. Luteolin promotes macrophage-mediated phagocytosis by inhibiting CD47 pyroglutamation. *Translational Oncology*. 2021;14(8):101129.
173. Hielscher-Michael S, Griehl C, Buchholz M, et al. Natural products from microalgae with potential against Alzheimer's disease: Sulfolipids are potent glutaminy cyclase inhibitors. *Marine Drugs*. 2016;14(11):203.
174. Kennedy LB, Salama AKS, A review of cancer immunotherapy toxicity. *CA: A Cancer Journal for Clinicians*. 2020; 70(2): 86–104.
175. Logtenberg MEW, Scheeren FA, Schumacher TN, The CD47-SIRPalpha immune checkpoint. *Immunity*. 2020;52(5):742–752.
176. Mantovani A, Longo DL, Macrophage checkpoint blockade in cancer – back to the future. *New England Journal of Medicine*. 2018;379(18):1777–1779.
177. van den Berg TK, Valerius T, Myeloid immune-checkpoint inhibition enters the clinical stage. *Nature Reviews Clinical Oncology*. 2019;16(5):275–276.
178. Gentles AJ, Newman AM, Liu CL, et al. The prognostic landscape of genes and infiltrating immune cells across human cancers. *Nature Medicine*. 2015;21(8):938–945.
179. Liu YE, Shi YF, Mitochondria as a target in cancer treatment. *MedComm*. 2020;1(2):129–139.
180. Wang X, Chen Y, Wang X, et al. Stem cell factor SOX2 confers ferroptosis resistance in lung cancer via upregulation of SLC7A11. *Cancer Research*. 2021;81(20):5217–5229.

Cite this article as: Liu Y, Shi Y & Wang P. Functions of glutaminy cyclase and its isoform in diseases. *Visualized Cancer Medicine*. 2023; 4, 1.